Copied from some of Benjy’s protocol. He derived it from manual, but without betamercapto so that we can do polyA on biotinylated RNA. Got about 2-3% as ng compared to total-RNA (2/27/14, Darach).

- Binding/wash buffer [20mM Tris-HCl (7.4), 500mM LiCl, 1mM EDTA]
- Low salt buffer [20mM Tris, 200mM LiCl, 1mM EDTA]
- Elution buffer [20mM Tris, 1mM EDTA]

1. Add 100ul of oligo dT beads (NEB) to eppendorfs, put in magnetic rack and discard the supernatant.
2. Wash beads once with 200ul binding buffer.
3. Add ~130ul binding buffer and ~20ul sample. This is flexible.
4. Vortex 20min RT.
5. Put in magnetic rack, remove (and keep probably) the supernatant. This is less polyA tails, likely a lot of ribosomal.
6. Resuspend in 200ul low salt buffer and vortex 3min.
7. Put in magnetic rack, remove (and keep probably) the supernatant (as above).
8. Resuspend beads in 20ul elution buffer, incubate 3min @ 50C.
9. Immediately put in magnetic rack and remove supernatant to new tube. This is your polyA-enriched fraction.