Non-denaturing RNA gel

Useful for confirming products, QC, and checking for RNA degradation.

Regarding RNA degradation, rinsing everything well with MilliQ water before using seems to work okay (for spike-in synth product checking). Just make sure you’re not using the gelbox that everyone uses for their minipreps (w/ RNaseA), and if you are dump that.

1. Pour a 1% agarose gel with 1x TAE and 5ul EtBr. Small combs suggested.
2. Mix ladder (ssRNA ladder) and samples each 50:50 with 2x RNA dye from NEB (comes with the ssRNA ladder). Put on 70C heatblock for 10 minutes to denature.
3. Put on wet ice for ~3min to cool before annealing.
4. Load gel. Run time of about 15min is bad for sizing products, but works for confirming products.